

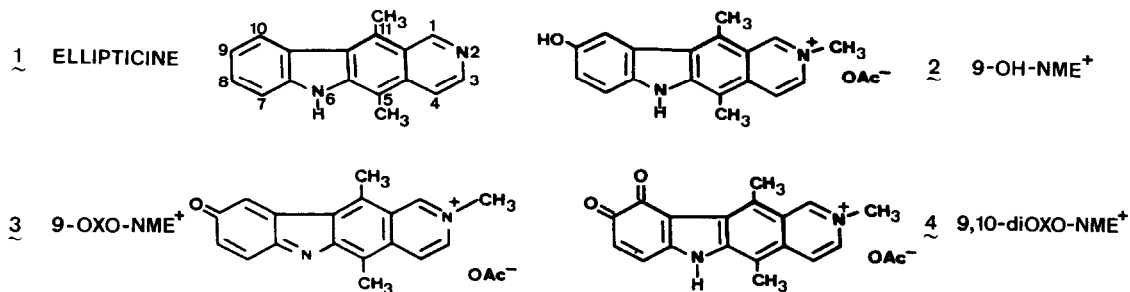
UNEXPECTED REGIOSPECIFIC ALKYLATION OF THE ANTITUMOR AGENT N²-METHYL-9-HYDROXYELLIPTICINIUM
 ACETATE WITH N, O OR S DONORS

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Summary : The quinone-imine derivative obtained by biochemical oxidation of [ ], N²-methyl-9-hydroxyellipticinium acetate, is found to react regiospecifically at C-10 with N, O and S nucleophiles.

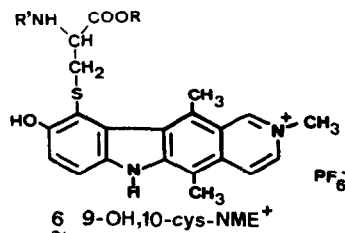
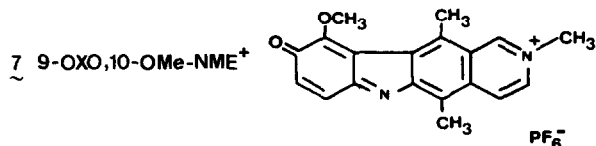
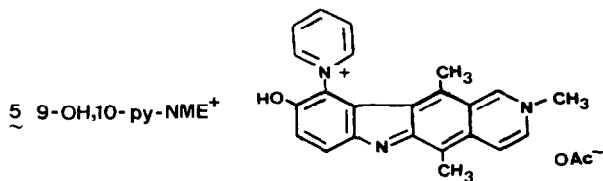
It has been reported¹ that some ellipticine derivatives manifest a much greater antitumour activity than ellipticine itself, ¹ (5, 11-dimethyl-6H-pyrido[4,3b] carbazole). One of these derivatives, ², N²-methyl-9-hydroxyellipticinium, (9-OH-NME⁺), which is obtained from ¹ by hydroxylation at C-9 and methylation of the pyridine nitrogen, shows a very interesting activity and is currently under clinical investigation².



The respective positions of the hydroxyl and the indolic (N-H) groups, which are para to each other, actually suggest that the drug ² might be activated *in vivo* through oxidative pathways. Our recent work³ on the formation of two quinone derivatives, ³ and ⁴, via peroxidase oxidation of ² by horseradish peroxidase (HRP) in presence of hydrogen peroxide, supports indeed this assumption.

Bearing in mind the alkylation of biological macromolecules by electrophilic quinone-imine derivatives and the implied consequences on biological activities^{4a}, we wish to report here our results about the addition of various nucleophiles (e.g. pyridine and cysteine derivatives) to ³, which occurs regiospecifically at C-10; we also want to report the formation of ⁷, the quinone-imine obtained by addition of a methoxy group at C-10 during the oxidation of ² by molecular oxygen in presence of Cu^I compounds. In both

cases H-10 only is substituted by nucleophiles to give a new adduct though, for steric reasons, this preference for H-10 over H-8 is rather unexpected.



6a R=H R'=H

6b R=Me R'=H

6c R=H R'=Ac

- Pyridine-adduct 5

This adduct is prepared by oxidation of 2, using HRP/H₂O₂ in a phosphate buffer 0.05 M (pH 7.8) containing 5 % (v:v) of pyridine. After 30 min the solution is concentrated and purified on a Servachrom XAD-2 resin column [eluent : methanol-ammonium acetate 0.1 M, 5:95(v:v), followed by methanol]. The yield is 38 %. No other adduct can be detected. Structure 5 agrees with the ¹H NMR and mass spectroscopy data of the compound obtained. ¹H NMR (90 MHz) of 5 in D₂O : CD₃COCD₃, 50:50(v:v). 1.92 (s, 3p, CH₃ COO⁻), 2.16 (s, 3p, Me₅), 2.85 (s, 3p, Me₁₁), 4.50 (s, 3p, N-Me), 7.53 (d, 1p, J = 8.5 Hz, H₇₍₈₎), 7.85 (d, 1p, J = 8.5 Hz, H₈₍₇₎), 8.41 (m, 2p, H₃, H₄), 8.57 (m, 2p, H'₃, H'₅ pyridine), 8.96 (m, 1p, H'₄ pyridine), 9.19 (m, 2p, H'₂, H'₆ pyridine) and 9.67 (s, 1p, H₁). The signals at 8.57, 8.96 and 9.1 could not be observed in the ¹H NMR spectrum of the pyridine-d₅ adduct. M = 413 (cation = 354). Observed peak in EI : 353 (358 for pyridine-d₅ adduct).

- Cysteine-adduct 6

To avoid a direct contact between the oxidant mixture and the -SH containing molecules, the coupling reaction is performed after 3 has been extracted with an organic solvent (methylene chloride) in presence of ammonium hexafluorophosphate. Cysteine derivatives are dissolved in methanol before the addition to 3 in CH₂Cl₂ is carried out. As cysteine adducts are rather instable when dissolved, the solutions were always kept at low temperature (-20 °C). The products are isolated after concentration and precipitation in a mixture acetone-ether-hexane. The yield is 30-40 %.

The ¹H NMR and mass spectroscopy data of the compounds obtained are in agreement with structures 6a, 6b and 6c, which all present a covalent C₁₀-S bond. The mass and ¹H NMR data of 6c are given here, as an example. Mass data (M = 583, cation = 438) ; FD : molecular peak at 439 (MH⁺) ; DCI (NH₃) : fragmentation peaks at 323 (ellip-S-CH₂), 309 (ellip-S) and 277.

¹H NMR (90 MHz) of 6c in D₂O : CD₃COCD₃ (3:1, v:v) : 1.63 (s, 3p, N-acetyl), 2.66 (s, 3p, Me₅)

3.19 (m, 2p -CH₂-cyst.), 3.37 (s, 3p, Me₁₁), 4.09 (m, 1p, -CH-cyst.), 4.51 (s, 3p, N⁺-Me) 7.25 (d, 1p, H₇ or H₈), 7.47 (d, 1p, J = 9.8 Hz, H₇ or H₈), 8.20 (broad s, 2p, H₃ and H₄) and 9.66 (s, 1p, H₁).

Other S-adducts have also been prepared, according to the same procedure.

- Methoxy-quinone-imine $\tilde{7}$

Hydroxylated molecules are usually readily oxidized by molecular oxygen in presence of Cu^I complexes^{5,6}. The oxidation of $\tilde{2}$, performed in methanol ($\tilde{2}$ is only poorly soluble in non-hydroxylated solvents) using molecular oxygen in presence of [(bipy)₂ Cu]Cl or [(bipy)₂ Cu] PF₆ is slow: The complete disappearance of $\tilde{2}$ is achieved after 24 hours only, while the same reaction performed with CuCl in methanol in presence of a small amount of pyridine leads to $\tilde{7}$ within one hour. After addition of NH₄PF₆ the solution is filtered twice on a Sephadex LH20 column [eluent: methanol]. After concentration of the solution, the product is precipitated by addition of ether. The yield is 75%. The structure $\tilde{7}$, a quinone-imine with a methoxy group at C-10, is compatible with IR, mass and ¹H NMR spectroscopy data. IR (CsBr): 1670 cm⁻¹ (C=O) and 1640 (C=N). Mass data (M=450, cation=305); EI: 306 (MH⁺); CI (NH₃): 307 (MH₂⁺).

¹H NMR in CD₃COCD₃: D₂O (2:1, v:v): 2.96 (s, 3p, Me₅), 3.24 (s, 3p, OMe), 3.34 (s, 3p, Me₁₁), 4.60 (s, 3p, N⁺-Me), 6.48 (d, 1p, J = 10.0 Hz, H₈), 7.91 (d, 1p, J = 10.0 Hz, H₇), 8.47 (2 d, 2p, J = 8.0 Hz, H₃, H₄) and 10.00 (s, 1p, H₁).

It is probable that the reaction proceeds through a nucleophilic attack of the methoxy group by a methoxy ligand of the copper complex, as described by Rogic et al. 6a,6b for the oxidation of diphenols by O₂/Cu^I in methanol.

The striking feature of the adduct derivatives described here is that they are all obtained from $\tilde{2}$ by a nucleophilic substitution at C-10. If the lack of reactivity of C-7 may be explained by the positive charge on N-2, which should favour the electrophilicity of positions 8 and 10 in $\tilde{3}$, the much lower reactivity of position 8 compared to 10 is more difficult to anticipate due to the sterical hindrance introduced at C-10 by the proximity of a methyl group at C-11. If $\tilde{3}$ were indeed formed as a metabolite, the N-acetyl-cysteine adduct $\tilde{6}$ should be found *in vivo* as the conjugate derived from the corresponding glutathione adduct. All these products have been tested *in vitro* on L1210 cells (Table 1) and have been found to be less cytotoxic than the starting compound $\tilde{2}$.

Table 1. Cytotoxicity of the adducts $\tilde{5}$, $\tilde{6}$ and $\tilde{7}$ (L1210 cells *in vitro*)

Compound	ID ₅₀ (48h)
9-OH-NME ⁺ $\tilde{2}$	0.075 μM
pyridine add. $\tilde{5}$	0.68 "
cysteine ester add. $\tilde{6b}$	0.79 "
cysteine acetyl add. $\tilde{6c}$	1.12 "
methoxy-10 cpd. $\tilde{7}$	3.0 "

ID₅₀ = concentration which inhibits the cell growth by 50% in 48 hours

The toxicity of compounds such as 2 might thus be related directly to their oxidability and therefore to their alkylation by biological nucleophiles following their oxidation *in vivo* [i.e. a "biooxidative alkylation" as opposed to the "bioreductive alkylation" proposed by Moore^{4b}]. Then the observed toxicity, which decreases from compound 5 to 7 can be explained as resulting from a nucleophilic addition on an oxidized form of 2 .

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References

1. G.M. SVOBODA, G.A. POORE and M.L. MONFORT, J. Pharm. Sci. 57, 1720 (1968).
2. a) C. PAOLETTI, J.B. LE PECQ, N. DAT-XUONG, P. JURET, H. GARNIER, J.L. AMIEL and J. ROUESSE, Recent Results in Cancer Research, 74, 107 (1980)
b) P. JURET, J.E. COUETTE, T. DELOZIER, J.Y. LETALAER, Bull. Cancer (Paris), 68, 224 (1981)
3. a) C. AUCLAIR and, C. PAOLETTI, J. Med. Chem. 24, 289 (1981).
b) G. MEUNIER, These de 3ème cycle, Université de Toulouse, June 1982.
4. a) S.D. NELSON, J. Med. Chem., 25, 753 (1982).
b) H.W. MOORE, Science, 197, 527, (1977).
5. M. MUNAKATA, S. NISHIBAYASHI and H. SAKAMOTO, J.C.S. Chem. Comm., 219 (1980).
6. a) T.R. DEMMIN, M.D. SWERDLOFF, and M.M. ROGIC, J. Amer. Chem. Soc., 103, 5795 (1981).
b) M.M. ROGIC and, T.R. DEMMIN, J. Amer. Chem. Soc., 100, 5472 (1978).
c) C. JALLABERT and, H. RIVIERE, Tetrahedron, 36, 1191 (1980).

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